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Review

BIOLOGICALLY ACTIVE COMPONENTS OF SOYBEANS AND SOY PROTEIN PRODUCTS - A REVIEW

Miroljub B. Barać, Slađana P. Stanojević and Mirjana B. Pešić

Soybeans provide a source of low-cost protein with good nutritional and physico-chemical properties. Recently, soybean has received much attention because of its potential role in preventing and treating several diseases, including cancer and other human chronic diseases. Health benefits of soy diet are attributed to the minor soybean constituents (called phytochemicals). Soybean contains a variety of phytochemicals with demonstrated anticancer activity, including bioactive proteins and polypeptides (trypsin inhibitors and the most recently discovered peptide lunasin), isoflavones, phytic acid, phytosterols and saponins. The present review provides an overview of recent knowledge about biologically active components of soybean.

KEY WORDS: Soybean; biologically active compounds; protein; health benefits

INTRODUCTION

Soybean is an abundant source of proteins that have long been recognized for high nutritional value and excellent physico-chemical properties in food. Also, soybean and soy products are rich sources of minor non-nutritive components with potential health benefits, in the literature often called phytochemicals.

Soybean contains many unique biologically active components including isoflavones, biologically active proteins and peptides, phytosterols, phytic acid and saponins. In traditional nutritional theory, many of these components have been considered as antinutrients. During the last two decades it has been found that they may exert beneficial health and therapeutic effects. The purpose of this work was to highlight recent knowledge about biologically active components of soybean, especially biologically active proteins and polypeptides.

Dr. Miroljub Barać, Assist. Prof., Slađana Stanojević, M.Sc., Assist., Mirjana B. Pešić, M.Sc., Assist., University of Belgrade, Faculty of Agriculture, 11080, Belgrade-Zemun, Nemanjina 6, Serbia and Montenegro

BIOLOGICALLY ACTIVE PROTEINS AND POLYPEPTIDES FROM SOYBEAN

Soybean is a good source of biologically active proteins and polypeptides, including protease inhibitors, lectins, low molecular weight polypeptides and the most recently discovered peptide lunasin. Recently, there has been an increased interest in their potential health benefits.

Protease inhibitors

Protease inhibitors, most commonly referred to as trypsin inhibitors, are the best known and the most studied biologically active proteins in soybean. Trypsin inhibitors are low molecular weight proteins that bind trypsin and interfere with protein hydrolysis during digestion (1). Soybean contains two major types of trypsin inhibitors: Bowman-Birk (BBI) and Kunitz inhibitors (KTI). Depending on the cultivar soybean has three Kunitz isoinhibitors and 5-12 Bowman-Birk inhibitors. Raw soybean contains 1.67% KTI and 0.4 % BBI (2). They are partially responsible for low digestibility of raw soybean. About 40% of the adverse effect of raw soy consumption is the result of their affects (3). Kunitz inhibitor (Figure 1a) consists of 181 amino acid residues with two disulfide bridges. One of them is essential for inhibitor activity. KTI stoichiometrically binds and inhibits only trypsin. The Bowman-Birk (Figure 1b) inhibitor has 70-80 amino acid residues with molecular weight of 7-8 kDa. The isoelectric point of BBI is 4.2 (5,6). This protein contains seven disulfide bridges and it is a rich source of amino acid residues with sulfur. BBI has two binding sites: a trypsin binding site (Lys^{16}-Ser^{17}) and a chymotrypsin reactive site (Leu^{43}-Ser^{44}). Thus, BBI can simultaneously bind both trypsin and chymotrypsin (7).

Trypsin inhibitors in raw soybean cause growth inhibition, pancreatic hypertrophy and hyperplasia in experimental animals (3). For a long time in the traditional nutritional theory trypsin inhibitors (TI) have been considered as typical antinutritional factors. Consequently, to reduce their activity, several treatments have been used. Thermal treatment is the most common method used for this purpose. The level of residual TI activity depends on several factors including treatment mode (level of temperature, time of heating), initial content of moisture, pH conditions and the presence of reducing agents (2, 8-12). It is well known that dry heat have no significant influence on TI activity, while moist heat treatments with steam at over the range of 0.5-2.0 bar (2, 8, 13, 14, 15), autoclaving (16) and steam jet cooking (17,18) were more effective. Additionally, the investigation conducted in our laboratory (19) showed that microwave roasting could be an effective method for the reduction of inhibitor activity. The type of inhibitor responsible for TI activity depends on treatment mode. Both types of inhibitor are responsible for the activity of microwave treated flour (19) and traditional soy protein concentrates (20). In opposite, our earlier investigation (2, 21) showed that the residual activity of moist heat treated flour is only the result of KTI.

In opposite to the traditional theory of nutrition, numerous investigations (22-29) have shown that protein inhibitors, especially BBI, had anticarcinogenic properties. Since 1992, BBI has achieved Investigational, New Drug status. BBI has been shown to be very effective in preventing carcinogenesis in the liver, lung, and gastrointestine in mouse models (22, 23). Kennedy and coworkers (24) conducted clinical trials to evaluate BBI as an anticarcinogenic agent in human populations. The mechanism of prevention is not clear. It has been suggested that protease inhibitors suppress both initiation and promotion stages of carcinogenesis. Due to the beneficial effects in nutrition, Kennedy and Szuhaj

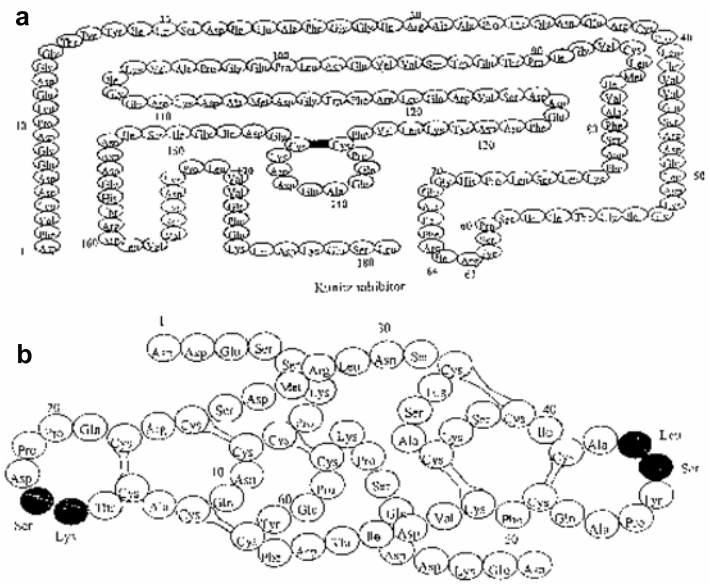

Fig. 1. Kunitz (a) and Bowman-Birk inhibitor (b) (4)

(30, 31) patented the isolation of Bowman-Birk concentrate from acidic aqueous extracts of hexane defatted soybeans. Also, Sessa and Wolf (32) prepared BBI concentrate from seed coats with considerably higher BBI activity than the Kennedy and Szuhaj patented concentrates.

Lectins

Lectins are a significant group of bioactive proteins found in almost all organisms, including plants, bacteria and viruses (33). Lectin originating from soybean seed is a tetrameric glycoprotein that accumulates during embryogenesis (34) and recognizes terminal α-linked 2-acetoamido-deoxy-D-galactosyl or α- or β—D-galactosyl sugar residues (35). Molecular weight of tetrameric form is 120 kDa. It normally constitutes 1-2 % of the seed protein mass. Lectins express ability to agglutinate red blood cells. It is a well recognized physiologic effect that is dependent on their specific high–affinity binding to particular carbohydrate moieties on the cell surface (36). SBA exists as multiple isolectins having similar binding and immunochemical properties. Native SBA consists of at least five isolectins: SBA-I, SBA-II and SBA-III (Figure 2).

The ingestion of pure lectins in the diet of animals has several biochemical, physiological, and nutritional implications such as agglutination of cells, stimulating pancreatic enzyme secretion (38-42). Once ingested lectin tends to stimulate intestinal cells, and thus can interfere with intestinal absorption of nutrients. Due to these facts lectin has been for a long time considered as antinutritient. In opposite, in the past 50 years it has been reported that plant lectins may have antitumor and anticarcinogenic activities that could be beneficial in cancer treatment (43-46). The exact mechanism(s) of the antitumor effect of plant lectins is not clear, although several have been proposed, including reduction of cell divi-

Fig. 2. The monomer of SBA showing the disposition of the tryptophan residues (37)

sion, increasing the number of macrophages, increasing the susceptibility of tumor cells to macrophage attack, serving as a bridge between tumor cells and macrophages, and improving the immunocompetence of tumor-bearing animals (34).

Soybean cultivars contain 6.5 g of lectin per kg of defatted soybean meal (47). Due to their protein structure, soy lectin is heat unstable and can be inactivated by different heat treatments. According to the heat stability soy lectins are similar to trypsin inhibitors and urease. Fasina and co-wokers (48) showed that the level or urease activity and TI activity can be reliable parameters of the degree of lectin activity reduction during soy meal autoclaving. Our earlier investigation (49) showed that moist heat treatments at 1.5-2.0 bar for 15 minutes completely destroy soy lectin polypeptides.

Bioactive peptides

Bioactive peptides may exist naturally or be derived from soy protein hydrolyzates. Usually, these peptides have common structural properties such as a relatively short peptide residue length (2-9 amino acid residues) and hydrophobic amino acid residues in addition to proline, lysine or arginine groups (50). Bioactive peptides are resistant to the action of peptidases (34, 51). These peptides act as physiological modulators during gastrointestinal digestion of soy products. Peptides derived from tryptic hydrolyzates of soybean proteins stimulate superoxide anions, which trigger nonspecific immune defense systems (52). Also, soybean peptides obtained from hydrolyzates express antioxidant activity (53), antiobesity effects (54). Furthermore, lunasin and hydrophobic peptides obtain from defatted proteins exhibit anticancer activity (50, 55).

Lunasin. Lunasin is the most recently discovered bioactive polypeptide originally isolated from soybean. While searching for methionine-rich proteins from midmaturation

soybean seed, researchers from the University of California, Berkley (56) isolated and cloned gene for small peptide termed as lunasin. More recently, lunasin has been isolated from barley (57). It is unique 43 amino acid peptide, whose carboxyl end contains nine residues of aspartic acid, an Arg-Gly-Asp cell adhesion motif, and a helix with structural homology to a conserved region of chromatin-binding proteins (58, 59). Now it is known that lunasin is a major component of Bowman-Birk protease inhibitor (34). Precisely, it is a linker peptide of BBI (60) with relative molecular mass of 5.45 ± 0.25 kDa detected by SDS-PAGE (52).

During seed development, lunasin peptide appears 5 weeks after flowering and persists in the mature seed. The content of lunasin in soybean seed varies with varieties and environment (51). Also, commercial soy protein products contain different amounts of lunasin. Soy isolates and hydrolyzed soy proteins contain the highest concentrations of lunasin. Soy protein concentrate, isolate, and hydrolyzate contain 2.81 ± 0.30, 3.75 ± 0.43, and 4.43 ± 0.59 g/100 g respectively, while soy flour and soy flakes contain 1.24 ± 0.22 g lunasin/100g (51). The content of lunasin detected in physiological phosphate buffer (PBS, O.1M pH 7.4) is shown in Table 1 (59).

Table 1. Lunasin content in some commercially available soy proteins (59)

Soy protein	**Lunasin (mg /g of protein)**	**Lunasin (% in product)**
Deffated soy flour	5.48 ± 0.17	0.054 ± 0.0017
Soy concentrate (alcohol washed)	8.72 ± 0.19	0.065 ± 0.0015
Soy isolate	6.92 ± 0.16	0.059 ± 0.0014
Soy concentrate (water washed)	16.52 ± 0.23	0.091 ± 0.0013

Synthetic lunasin is heat stable, surviving temperatures up to 100°C for 10 minutes (34).

According to Galvez et al. (58) transfection of mammalian cells with the lunasin gene leads to mitotic arrest and cell death characterized by cell lysis and chromosome fragmentation. This effect is attributed to binding of its poly-aspartyl carboxyl end to regions of hypoacetylated chromatin like that found in centromeres (59).

Lunasin is a promising chemopreventive peptide from soybean. Galvez et al (55) reported *in vitro* and *in vivo* chemopreventive properties of chemically synthesized lunasin against chemical carcinogenesis. This may be a result of the RGD-cell adhesion motif and chromatin-binding properties (60). Also, animal studies showed that lunasin inhibited skin tumorigenesis in a mouse skin cancer model when applied topically (55). In all of these experiments synthetic peptide was used. Due to the high cost of synthetic lunasin, there is a need to isolate, characterize and demonstrate the biological activity of natural polypeptide from soybean.

Hydrophobic peptides. During processing of soy protein products, especially during fermentation or heat treatment, many peptides can be formed. Some of these peptides

exert various physiological activities (61). Kim et al. (62) reported that low molecular weight peptides derived from soybean proteins exhibited cytotoxicities on several cell tumor lines. Kitts et al. (52) isolated and characterized nonapeptide with cancer reducing activity of thermoase-hydrolyzed defatted soybean. Anticancer peptide was extracted with ethanol. The molecular weight of this peptide was 1157 Da and the amino acid sequence was X-Met-Leu-Pro-Ser-Tye-Ser-Pro-Tyr.

NON-PROTEIN BIOACTIVE COMPONENTS FROM SOYBEAN

Phytic acid. Phytic acid (PA, myo-inositol 1,2,3,4,5,6-hexakis-dihydrogen phosphate, Figure 3) is the storage form of phosphorus in soybean seeds. Soybean seed and soy protein products have relatively high content of phytic acid. The phytic acid content of soy protein products is determined by the conditions employed for its preparation. Commercial soy protein products contain 1.4-3% PA (64), while the content of the laboratory prepared isolates and concentrates are 2.89% and 4.81-4.95 %, respectively (64, 20). The major portion of phytate is in the 7S protein fraction, while glycinin (11 S-protein) contains only about 0.07% phytate (65).

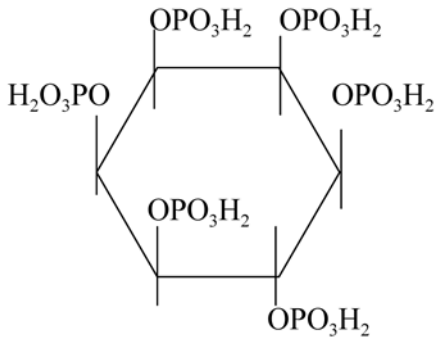

Fig. 3. Phytic acid (63)

Phytic acid can interact with minerals, proteins and starch. The degree of interaction of phytic acid and proteins is affected by the protein charge, conformation and ionic strength of the solution at a given pH. The interaction of PA with proteins and minerals is shown in Figure 4.

Due to the abillity of chelating of minerals and binding of proteins and starch PA can reduce their digestibility (66). Consequently, it has been considered as an antinutritient. On the contrary, it is well known today that low concentration of PA has some beneficial effect in nutrition, which include controling dental caries, improving oxygen–providing ability of red blood cells and cancer preventing activity, preventing of cardiovascular diseases and diabetes (1,67,68). According to Wang and Wixon (1), anticarcinogenic effect of phytic might be result of their interaction with starch, protein and minerals.

Soy isoflavones. Soybeans and soy protein products are abundant source of isoflavones. Isoflavones are class of phenolic components. The major isoflavones of soybeans are daidzein, glycitein, and genistein. Each of them is found in four chemical forms: the unconjugated form or aglycone; the conjugated form or glucoside (daidyin, genistin, and glycitin); acetylglucoside; and malonylglucoside (69,70,). Structures of soy isoflavone aglucones are shown in Figure 5.

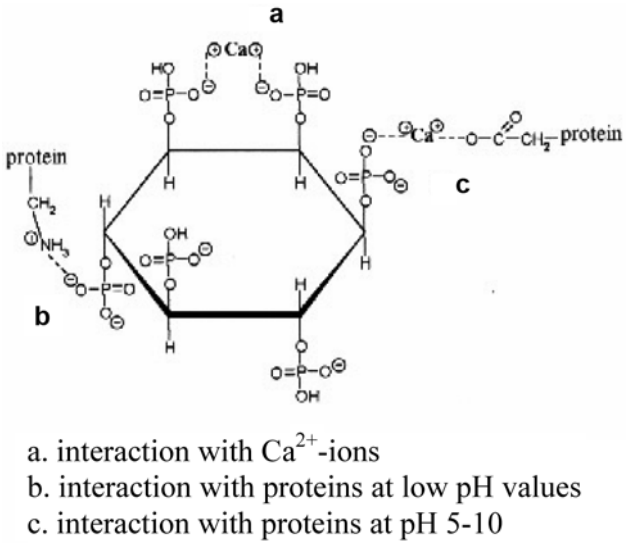

Fig. 4. The interaction of PA with proteins and Ca-ions (62)

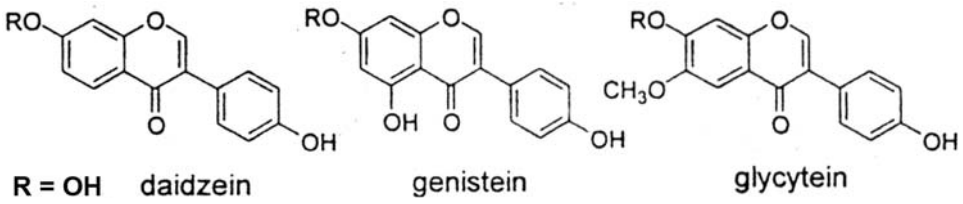

Fig. 5. Structure of soy isoflavone aglucones (69)

The concentration of isoflavones in soybean seed and soy ingredients significantly varies due to genetic variation of soybean cultivars, environment conditions where the soybean is rised, and processing conditions used for soy ingredient preparation. For example, only 23% of the total isoflavones in soy flour remaind after the soy flour had been processed to soy protein isolates (1). According to Kurzer and Xu (71), soybean-based products contain approximatly 0.2-1.6 mg of isoflavones per g dry weight.

Isoflavones have multi-biological and pharmacological effects in animals and humans including estrogenic and antiestrogenic effects (72, 73), cell signalling conduction (74-76), as well as cell growth and death. Soy isoflavones have been associated with reduced incindences of breast, prostate and lung cancer (77-79) cardiovascular diseases (80) and osteoporosis (77, 81). The mechanism through which isoflavones may exert beneficial effects are not only based on their estrogenic properties, but also on their role as protein tyrosin kinase inhibitors, as regulators of gene transcription, antioxidants, as well as by altering some enzyme activities (70).

Phytosterols. Soybeans are rich source of phytosterols. The total content of these components is 0.3-0.6 mg/g. β-sitosterols, campesterol and stigmasterol (Figure 6) are

three major phytosterols in soybean. Although these components are structurally related to cholesterol, they have many "anticholesterol" properties (82). Phytosterols decrease absorption of cholesterol due to the inhibition of bile cholesterol micelle formation and competition for the enzyme required for cholesterol uptake. Also, *in vitro* animal studies have demonstrated that these components have anticarcinogenic properties. Namely, phytosterols competitively inhibit cholesterol dehydrogenase and other bacterial enzymes in the colon. Consequently, they reduce the production of the secondary biles that may cause colon cancer (83).

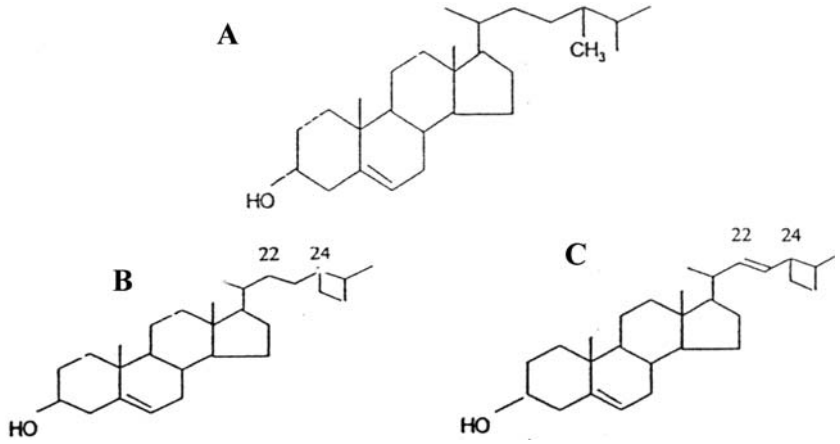

Fig. 6. Structures of three major phytosterols in soybeans (82)
A.campesterol, B.sitosterol, C.stigmasterol

Saponins. Saponins are a group of heat-stable glycosides of triterpenoids or steroids. Soybeans are rich source of saponins. Also, saponins are present in all of the soy protein products except for those that are extracted with alcohol. Soybean saponins are classified into three groups A, B, C. Group A saponins, known as bis-desmosides contain two ether-linked shugar chains, while groups B and C, known as mono-desmosides, contain one shugar chain. There are three dominant forms of saponins in raw soybean: soyasaponin ag, soyasaponin βg, and soyasaponin βa, in concentrations ranging from 1 to 3 mg/g (1). Due to the presence of triterpene or steroid part of their molecule, which is hydrophobic, and the presence of hydrophilic sugar chains, these compounds are amphiphilic. As a result of surface characteristics, saponins have hemolytic, hypocholesterolemic, immunostimulatory and antitumor activities (84, 85).

CONCLUSION

Soybeans are not only excellent sources of high-quality vegetable proteins but also contain appreciable amounts of biologically active components. Soybeans have many unique biologically active components including trypsin inhibitors, lectins, isoflavones, phytosterols, saponins, phytic acid and the most recently discovered peptide called lu-

nasin. For a long time, many of these components have been considered as antinutrients. However, the current literature contains numerous reports that most of these components exert beneficial health and therapeutic effects at low concentrations. Due to their role in preventing and treating several diseases, including cancer and other chronic diseases, with the careful selection of processing conditions soy-based products may become the "superfoods" of the millennium.

REFERENCES

1. Wang, C. and R. Wixon: Phytochemicals in soybeans – Their potential health benefits. Inform **10** (1999) 315-320.
2. Veličković, D.,Vucelić–Radović, B., Barać, M., D. Simić: Change of Trypsin Inhibitor Activity as a Function of Pressure and the Duration of Thermal Treatment of Soybean Flour. Rev. of Res. Work Fac. Agr. Belgrade **37** (1992) 109-116.
3. Liener, I. E.: Factor Affecting the Nutritional Quality of Soya Products. J. Am. Oil Chem. Soc. **49** (1981) 406-415.
4. Koide, T. and T. Ikenaka: Studies on Soybean Trypsin-Inhibitors 3. Amino acid Sequence of the Carboxyl-Terminal Region and the Complete Amino Acid Sequence of the Soybean Trypsin Inhibitor (Kunitz). Eur. J. Biochem. **32** (1973) 417-431.
5. Birk, Y.: Protein proteinase inhibitors in legume seed - Overview. Arch. Latinoam. Nutr. **44** (1996) 26S-30S.
6. Birk, Y., Gertler, A. and S. Khalef: A Pure Trypsin Inhibitor from Soya Beans. Biochem. J. **87** (1963) 281-284.
7. Birk, Y.: The Bowman-Birk Inhibitor. Int. J. Peptide Protein Res. **25** (1985) 113-119.
8. Veličković, D., Vucelić–Radović, B., Barać, M. and S. Stanojević,: Effect of Soybean Thermal Inactivation on Trypsin Inhibitor Activity of Protein Isolate. Rev. of Res. Work Fac. Agr. Belgrade **42** (1997) 229-235.
9. Veličković, D.,Vucelić–Radović, B., Barać, M., Stanojević, S. and D. Simić: The Effect of Pressure and Duration of Heating on Inactivation and Protein Composition of Soy Flour. 2nd Yugoslav Congress of Food Technology, Belgrade, September 1995, Book of Abstracts, p. 82.
10. Veličković, D., Vucelić–Radović, B., Barać, M., S.Stanojević: Uporedno ispitivanje aktivnosti inhibitora tripsina i ureaze u različito tretiranim frakcijama lomljenog sojinog zrna, III Jugoslovenski simpozijum prehrambene tehnologije, Beograd, Zbornik radova, sveska **III** (1998) 44-48.
11. Sessa, D.J. and J.A. Bietz: Toasted Soybean Flour Components with Trypsin inhibitor activity. J. Am. Oil Chem. Soc. **63** (1986) 784-788.
12. Friedman, M., Gumbman, M. R., Brandon, D. L., A. H. Bates: Inactivate and analysis of soybean inhibitors of digestive enzymes, in Food Proteins. Eds. J.E. Kinsela and W.G. Soucie, A.O.C.S., Champaign IL (1989) pp 296-342.
13. Friedman, M. and D.Brandon: Nutritional and Health Benefits of Soy Proteins. J. Agric. Food Chem. **49** (2001) 1069-1086.
14. Veličković, D., Vucelić–Radović, B., Barać, M., and S. Stanojević: Change of Soybean Polypeptide Composition during Thermal Inactivation of Trypsin Inhibitors. Acta Periodica Technologica **31** (2000) 193-199.

15. Vucelić-Radović, B., Barać, M., Stanojević, S. and M. Pešić: Biologically Active Components of Soy Protein Isolates Prepared from Hydrothemally Treated Cracked Soybeans. Journal of Scientific Agricultural Research **64** 1 (2003) 3-19.
16. Stanojević, S., Vucelić-Radović, B., Barać, M., M. Pešić: The Effect of Autoclaving on Soluble Protein Composition and Trypsin Inhibitor Activity of Cracked Soybeans. Acta Periodica Technologica **35** (2004) 49-58.
17. Johnson, L. A., Deyoue, C. W., Hoower, W.J. and J. R. Shwenke: Inactivation of Trypsin Inhibitors in Aqueous Extracts of Soybean by Direct Steam Infusion. Cereal Chem. **57** (1980) 376-393.
18. Wang, C., and Johnson, L. A.: Functional Properties of Hydrothermally Cooked Soy Protein Products. J. Am. Oil Chem. Soc. **78** (2001) 189-195.
19. Barac, M. and S. Stanojevic: The Effect of Microwave Roasting on Soybean Protein Composition and Components with Trypsin Inhibitor Activity. Acta Alimentaria **34** (2005) 23-31.
20. Barać, M.: Chemical and enzymatic modification of soy protein concentrates, University of Belgrade, 2002.
21. Barać, M.: The Change of 7S, 11S Proteins and Trypsin Inhibitor Activity During Thermal Inactivation of Soy Flour, University of Belgrade, 1993.
22. Kennedy, A.R.: The Bowman–Birk Inhibitor from Soybeans as an Anticarcinogenic Agent. Am. J. Clin. Nutr. **68** (1998a) 1406S–1412S.
23. Kennedy,A.R.:Chemopreventiveagents:proteaseinhibitors.Pharmacol.Ther.**78**(1998) 167–209.
24. Kennedy, A.R.: Cancer Prevention by Bowman–Birk inhibitor concentrate (BBIC), in Cancer Nutrition Eds. Prasad, K.N. and W.C. Cole, IOS Press, Amsterdam (1998) pp. 93–97.
25. Zhang, L., Wan, X.S., Donahue, J.J., Ware, J.H. and A.R. Kennedy: Effects of the Bowman–Birk Inhibitor on Clonogenic Survival and Cisplatin- or Radiation-Induced Cytotoxicity in Human Breast, Cervical, and Head and Neck Cancer Cells. Nutr. Cancer **33** (1999) 165–173.
26. Wan, H.S., Ware, J.H., Zhang, L., Newberne, P.M., Evans, S.M., Clark, L.C. and A. R. Kennedy: Treatment with Soybean-Derived Bowman-Birk Inhibitor Increases Serum Prostate-Specific Antigen Concentration while Suppressing Growth of Human Prostate Cancer Xeno-graphs in Nude Mice. Prostate **4** (1999) 243-248.
27. Billings, P.C., St.Clair, W.H., Maki, P.A. and A.R. Kennedy: Distribution of the Bowman-Birk Inhibitor in Mice Following Oral Administration. Cancer Lett. **62** (1992) 191-197.
28. Billings, P.S., Brandon, D.L. and J.M. Habre: Internalization of the Bowman-Birk Protease Inhibitor by Intestinal Epithelial Cells. Eur. J. Cancer **27** (1991) 903-908.
29. Armstrong, W.B., Kennedy, A.R., Atiba, J., McLaren, C.E. and F.L. Meyskens: Single-dose Administration of Bowman-Birk Inhibitor Concentrate in Patients with Oral Leukoplakia. Cancer Epidemiol. Biomarkers Prev. **9** (2000) 43-47.
30. Kennedy, A.R. and B.F. Szuhaj: US Patent No. **5**, 217 (1993) Method of Making Soybean Bowman–Birk Inhibitor Concentrate and Use of Same as a Human Cancer Preventative and Therapy, 717.

31. Kennedy A.R. and B.F. Szuhaj: US Pat. No. **5**, 338 (1994), Bowman–Birk Inhibitor Product for Use as an Anticarcinogenesis Agent, 547.
32. Sessa, D.J. and W.J. Wolf: Bowman-Birk Inhibitors in Soybean Seed Coats. Ind. Crops and Products **14** (2001) 73-78.
33. Sharon, N. and H. Lis: Lectins. Proteins with Sweet Tooth: Function in Cell Recognition. Essays Biochem. **30** (1995) 59-75.
34. Gonzales de Mejla, E., Bradford, T. and C. Hasler: The Anticarcinogenic Potential of Soybean Lectin and Lunasin. Nutrit. Rev. **61** (2003) 239-246.
35. Su, L.C., Pueppke, S.G. and H.P. Friedman: Lectins and Soybean Rhizobium Symbiosis. I. Immunological Investigations of Soybean Lines, the Seeds of which have been reported to Lack the 120.000 Dalton Soybean Lectin. Biochim.Biophys. Acta **29** (1980) 292-304.
36. Rabijns, A., Verboven, C. and P. Rouge: Structure of Legume Lectin from Bark of Robinia and its Complex with N-acetylgalactosamine. Proteins **44** (2001) 470-480.
37. Sharmistha, S. and S. Avadhesha: Oligomerization Endows Enormous Stability to Soybean Agglutinin: A Comparison of the Stability of Monomer and Tetramer of Soybean Agglutinin. Biophys. J. BioFAST (2005).
38. Pusztai, A., Grant, G., Bardocz, S., Gelericser, E. and G. Hajos: Novel Dietary Strategy for overcoming Nutritional Effects of Soybean Whey of High Agglutinin Content. Br. J. Nutr. **77** (1997) 933-945.
39. Grant, G., Alonso, R., Edwards, J.E. and S. Murray: Dietary Soya Beans and Kidney Beans Stimulate Secretion of Cholecystokinin and Pancreatic Digestive Enzymes in 400-day-old Hooded-wistar Rats but Only Soya Beans Induce Growth of the Pancreas. Pancreas **20** (2000) 305-312.
40. Grant, G., Henderson, L.T., Edwards, J.E., Ewan, E.C., Bardocz, S. and A. Pusztai: Kidney Bean and Soybean Lectins Cause Enzyme Secretion by Pancreatic Acini in Vitro, Life Sci. **60** (1997) 1589-1595.
41. Lalles, J.P., Tukur, H.M., Toullec,R. and B.G. Miller: Analytical Criteria for Predicting Apparent Digestibility of Soybean Protein in Preruminant Calves. J. Dairy Sci. **79** (1996) 475-482.
42. Douglas, M.W., Persons, G.M. and T. Humowitz: Nutritional Evaluation of Lectin-Free Soybeans for Poultry. Poultry Sci. **78** (1999) 91-95.
43. Pryme, I.F.: Bardoczs Anti-Cancer Therapy: Diversion of Polyamines in the Gut. Eur J Gastroenterol Hepatol. **13** (2001) 1041-1046.
44. Evans, R.C., Fear, S. and D. Ashby: Diet and Colorectal Cancer: An Investigation of the Lectin/Galactose Hypothesis. Gastroenterology **122** (2002) 1784-1792.
45. Ganguly, C. and S. Das: Plant Lectins as Inhibitors of Tumor Growth and Modulators of Host Immune Response. Chemotherapy **40** (1994) 272-278.
46. Liener, I.E.: From Soybean to Lectins: a Trail of Research Revisited. Carbohydr. Res. **21** (1991) 1-5.
47. Vasconcelos, I.M., Siebra, E.A. and A.A.B. Maia: Composition, Toxic, and Anti-nutritional Factors of Newly Developed Cultivars of Brazilian Soybean (Glycine max). J. Food Sci. Agric. **75** (1997) 419-426.

48. Fasina, Y.O., Classen, H.L., Garlich, J.D., Swaisgood, H.E. and D.A. Clare: Investigating the Possibility of Monitoring Lectin Levels in Commercial Soybean Meals Intended for Poultry Feeding Using Steam-heated Soybean Meal as a Model. Poultry Science **82** (2003) 648-656.
49. Veličković, D., Vucelić–Radović, B., Barać, M., S. Stanojević: Uticaj pritiska vodene pare i dužine trajanja termičkog tretmana na inaktivaciju i proteinski sastav sojinog brašna, u Savremeni trendovi u prehrambenoj tehnologiji, ur. D.B. Obradović i M.A. Janković, Poljoprivredni fakultet, Beograd (1995) 226-239.
50. Kim, S.E., Kim, H.H., Kim, J.Y., Kang, Y.I., Woo, H.J., and H.J. Lee: Anticancer Activity of Hydrophobic Peptides from Soy Proteins. BioFactors **12**, 1 (2000) 151-155.
51. De Mejla, G., Vasconez, M., de Lumen, B.O. and N. Randal: Lunasin Concentration in Different Soybean Genotypes, Commercial Soy Protein and Isoflavone Products. J. Agric. Food Chem. **52** (2004) 5882-5887.
52. Kitts, D.D. and K. Weiler: Bioactive Proteins and Peptides from Food Sources: Applications of Bioprocesses Used in Isolation and Recovery. Curr. Pharm. Des. **9,** 16 (2003) 1309-1323.
53. Pena-Ramos, E. A. and Y. L. Xiong: Antioxidant Activity of Soy Protein Hydrolysates in a Liposomal System. J. Food Sci. **67** (2002) 2952-2956.
54. Nakamori, T.: Antiobesity Effects of Soy Proteins and Soy Peptides. Food Style **21**, 5 **(**2002) 86-88.
55. Galvez, A., Chen, N., Macasieb, J. and B.O. de Lumen: Chemopreventive Property of a Soybean Peptide (Lunasin) that Binds to Deacetylated Histones and Inhibits Acetylation. Cancer Res. **61** (2001) 7473-7478.
56. Galvez, A.F., Revilleza, M.JR, and B.O. de Lumen: A Novel Methionine-rich Protein from Soybean Cotyledon: Cloning and Characterization of cDNA (accession No. AF005030). Plant Gene Register #PGR97-103. Plant Physiol. **114** (1997) 1567-1569.
57. Jeong, H.J., Lam, Y. and B.O. de Lumen: Barley Lunasin Suppresses Ras-induced Colony Formation and Inhibits Core Histone Acetylation in Mammalian Cells. J. Agric. Food Chem. **50** (2002) 5903-5908.
58. Galvez, A.F. and B.O. de Lumen: A Soybean cDNA encoding a Chromatin-binding Peptide Inhibits Mitosis of Mammalian Cells, Nat. Biotechnol. **17** (1999) 495-500
59. Jeong, H.J., Park, H.J., Lam, Y. and B.O. de Lumen: Characterization of Lunasin Isolated from Soybean. J. Agric. Food Chem. **51** (2003) 7901-7906.
60. Akiyama, T., Ishida, J., Nakagawa, S., Watanabe, S., Itoh, N., Shibuya, M. and Y. Fukami: Genistein, a Apecific Inhibitor of Tyrosine-specific Protein Kinase. J. Biol. Chem. **262** (1995) 5592-5595.
61. Clare Mills, E.N., Aleocer, M.J.C. and M.R.A. Morgan: Biochemical Interaction of Food Derived Peptides. Trends Food Sci. Technol. **3** (1992) 64-68.
62. Kim, J. Y., Woo, H.J., Ahn, C.W., Nam, Z.I., H.J. Lee: Cytotoxic Effect of Peptides Fractionated From Bromelain Hydrolizates of Soybean Proteins, Food Sci. Biotech. **8** (1999) 333-227.
63. Thomson, L: Reduction of Phytic Acid Concentration in Protein Isolates by Acilation Techniques, J. Am. Oil. Chem. Soc. **64** (1987) 1712-1719.

64. Johnson, D. W. and S. Kikuchi,: Proceedings of the World Congress on Vegetable Protein Utilization in Human Foods and Animal Food Stuffs, Applewhite, T.H.,AOCS, Champain USA, (1989) 73-78.
65. Veličković, D., Vucelić–Radović, B., Blagojević, S., Barać, M., Stanojević, S., M. Ljubičić: A Modification of a Method for Phytic Acid Determination. J. Serb. Chem. Soc. **64** (1999) 303.
66. Morr, C.V.: Current Status of Soy Protein Functionality in Food Systems. J. Am. Oil Chem. Soc. **67** (1990) 265-272.
67. Thompson, L.U. and L. Zhang: Phytic Acid and Minerals: Effect on Early Markers of Risk for Mammary and Colon Carcinogenesis. Carcinogenesis **12** (1991) 2041-2047.
68. Graf, E. and J.W. Eaton: Suppresion of Colonic Cancer by Dietary Phytic Acid. Nutr. Cancer. **19**, 1 (1993) 11-16.
69. Ren, M.K., Kuhn, G., Wegner, K. and J. Chen: Isoflavones, Substances with Multi-Biological and Clinical Properties. Eur. J. Nutr. **40** (2001) 135-146.
70. Kudou, S., Fleuru, Y., Welti, A., Mognolato, D., Uchida, T., Kitamura, K. and K. Okubo: Malonyl Isoflavone Glucosides in Soybean Seeds (Glycine Max Merrill). Agric. Biol. Chem. **55** (1991) 2227-2233.
71. Kurzer, M.S. and X. Xu: Dietary Phytoestrogenes. Ann. Rev. Nutr. **17** (1997) 353-381.
72. Setchell, K.D.: Phytoestrogens: the Biochemistry, Physiology, and Implicatios for Human Health of Soy Isoflavones. Am. J. Clin. Nutr. **68** (1998) 1333S-1346S.
73. Setchell, K.D. and A. Cassidy: Dietary Isoflavones: Biological Effects and Relevance to Human Health. J. Nutr. **129** (1999) 758S-767S.
74. Dalu, A. Haskel, J.F., Coward, L and C.A. Lamartiniere: Genistein, A Component of Soy, Inhibits the Expression of the EGF and ERBB2/Neu Receptors in the Rat Dorsolateral Prostate. Prostate **37** (1998) 36-43.
75. Agarwall, R.: Cell Signaling and Regulators of Cell Cyrcle as Molecular Targets for Prostate Cance Prevention by Dietary Agents. Biochem. Pharmacol. **l60** (2000) 1051-1059.
76. Zhang, Z.H. and Q. Wang: Modulation of a Cloned Human A-Type Voltage-Gated Potassium Channel by the Protein Kinase Inhibitor Genistein, Pflugers Arch. **440** (2000) 784-792.
77. Murkies, A.L., Wilcox, G. and S.R. Davis: Clinical Review. Phytoestrogenes. J. Clin. Endocrinol. Metab. **83** (1998) 297-303.
78. Anderson, J.J.B., Anthony, M., Messina, M. and S. Garner: Effects of Phyto-oestrogens on Tissue, Nutr. Res. Rev. **12** (1999) 75-116.
79. Adlercreutz, H., Mazur, W., Bartles, P., Watanabe, S., Lundin, E., Bergh, A., Damber, J.E., Aman, P., Widmark, A., Johansson, A., Zhang, J.X. and G. Hallmans: Phytoestrogens and Prostate Disease, J. Nutr. **130** (2000) 648-659.
80. Anthony, M.S.: Soy And Cardiovascular Disease: Cholesterol Lowering and Beyond, J. Nutr. **130** (2000) 662S-663S.
81. Davis, S.R., Dalais, F.S., Simpson, E.R., A.L. Murkies: Phytoestrogens in health and disease, Recent Prog. Horm. Res. **54** (1999) 185-210.

82. Carroll, K.K. and E.M. Kurowska: Soy Consumption and Cholesterol Reduction: Review of Animal and Human Studies, J. Nutr. **125** (1995) 594S-597S.
83. Rao, A.V. and S.A. Janezic: The role of dietary phytosterols in colon carcinogenesis, Nutr. Cancer **18** (1992) 43- 52.
84. Sidhu,G.S. and D.G. Oakenfull: A mechanism for the hypocholesterolemic activity of saponins, Br. J. Nutr. **55** (1986) 643-649.
85. Rao, A.V. and M.K. Sung: Saponins as anticarcinogens, J. Agric. Food Chem. **125** (1995) 717S-724S.

БИОЛОШКИ АКТИВНЕ КОМПОНЕНТЕ ЗРНА СОЈЕ И ПРОТЕИНСКИХ ПРОИЗВОДА ОД СОЈЕ

Мирољуб Б. Барађ, Слађана П. Станојевић и Мирјана Б. Пешић

Соја представља добар извор нутритивно високовредних протеина погодних физичко-хемијских особина. Данас се соји поклања велика пажња као потенцијално значајном извору компоненти које могу имати улогу у превенцији и лечењу неких хроничних и тешких болести, као што су болести срца и различити облици канцера. Позитивни ефекти исхране сојом приписују се мање заступљеним биолошки активним компонентама сојиног семена. Ове супстанце у литератури се често називају и «фитохемикалијама». Соја садржи цео спектар биолошки активних супстанци које испољавају антиканцерогена својства попут биоактивних протеина и полипептида (инхибитори протеаза, лектин и најновије детектовани полипептид - лунасин), фитинска киселина, изофлавони, фитостероли и сапонини. Овај рад даје преглед данашњих сазнања о биолошки активним компонентама зрна соје и протеинских производа од соје.

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